

## Human zygote morphological indicators of higher rate of arrest at the first cleavage stage

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### Summary

A little studied aspect of developmental arrest (DA) in ART is zygote arrest (ZA). Etiologically, blockage at the first cleavage stage includes molecular and chromosomal anomalies, some of which manifest morphologically. Given considerations on embryo culture, transfer and cryopreservation, optimal zygote selection is very important. The aim of this study was to ascertain whether zygote morphological features were indicators of increased ZA. In this study we performed a prospective, observational study of 2105 zygotes obtained from consecutive patients who were undergoing IVF/ICSI treatment, of which 43 (2%) suffered ZA. Morphological features observed under the inverted microscope were qualitatively categorized: pronuclear size, nucleolar precursor bodies (NPB) alignment, light and dark halos, polar body placement and fragmentation observed at 16–18 h post-insemination. We compared these features in blocked versus cleaved zygotes at 48 h and found significant correlations ( $p < 0.05$ ) between ZA and three features: the absence of a light halo ( $p = 0.001$ ), the absence of a dark halo ( $p < 0.005$ ), and non-aligned NPB ( $p < 0.05$ ). We can say that certain morphological features are indicators of significantly increased zygote arrest. These findings may be of utility for optimal zygote selection and culture strategies, especially in countries under restrictive conditions.

Keywords: First cleavage stage, Dark halo, Light halo, Morphological features, Nucleolar precursor body alignment, Zygote arrest

### Introduction

During *in vitro* culture from oocytes to embryos, approximately 50% of developmental arrest (DA) occurs in the first week (Hardy *et al.*, 2001; Racowsky, 2002). This study focuses on one aspect of the phenomenon: zygote arrest (ZA) observed at the first cleavage stage, which marks the transition from zygote to embryo.

The known causes of ZA include suboptimal culture conditions and inadequate oocyte maturation (Favetta, 2007). Apoptosis also plays a determinant role in DA (Hardy *et al.*, 2001). A high proportion of chromosomal abnormalities have been found in blocked zygotes (Benkhalifa *et al.*, 2003), although this finding was also related with some degree of cytoplasmic or nuclear immaturity.

The human zygote inherits its mitotic potential from the male gamete. In most species, (the mouse is a notable exception), the centrosome is paternally inherited (Palermo *et al.*, 1994; Sathanathan *et al.*, 1996). However, other factors influence this process. Zygote arrest 1 (*Zar 1*) is a maternal-effect gene critical for the oocyte-to-embryo transition (Wu *et al.*, 2003) and thus participates in the initiation of embryo development and the control of fertility in mammals.

Other factors may play an important role; Rawe *et al.* (2003) suggested that abnormal molecular

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nucleo-cytoplasmic trafficking might accompany human fertilization arrest.

In addition to genetic and molecular factors, dysmorphic zygotes with pronuclear size differences of more than 4  $\mu\text{m}$  show significantly increased DA (Sadowy *et al.*, 1998). The notion that there may be other easily observable morphological aspects related to initial DA prompted this study. Although the rate of ZA at the cleavage stage is low (circa. 2%), a rapid, simple method of early ZA prediction would benefit any assisted reproduction centre for embryo selection at the initial stages.

## Material and methods

We performed a prospective observational study of 2105 zygotes at 16–18 h post-insemination, using an inverted microscope with magnification of  $\times 10$ – $20$ – $40$  (Diaphot; Nikon Corporation). The zygotes were obtained from consecutive patients undergoing IVF or ICSI treatment at our Human Reproduction Unit.

### Ovarian stimulation

Pituitary suppression was achieved using a long gonadotrophin-releasing hormone analogue (GnRHa, Decapeptyl; Ipsen) protocol, followed by ovarian stimulation with recombinant follicle stimulating hormone (Gonal; Serono Laboratories). Transvaginal ultrasound guided follicle puncture took place 34 to 36 h after human chorionic gonadotrophin injection.

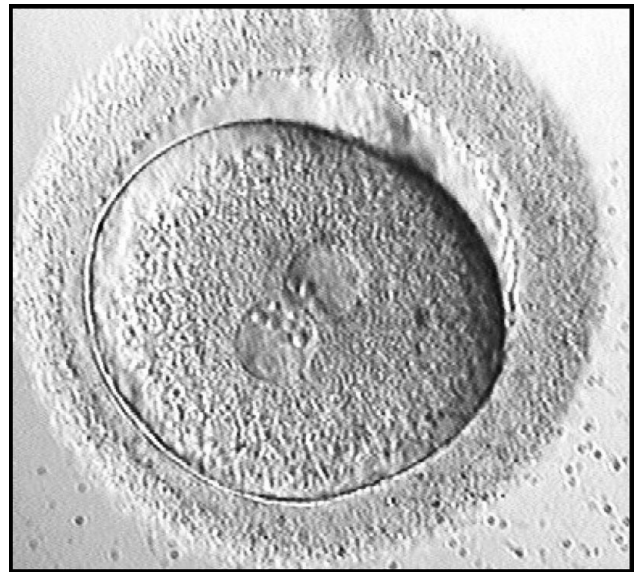
### IVF/ICSI procedures

For IVF procedure, the oocytes were cultured at 6%  $\text{CO}_2$  and 37°C for at least 3 h and after that time placed in a plate with macrodrops (100  $\mu\text{l}$ ) of a suspension of IVF (Medicult) with the spermatozoa (300,000 to 500,000 spermatozoa/ml).

For the ICSI procedure, the cumulus-corona cells were initially removed by exposure to a flushing medium (Medicult) with 80 IU/ml of hyaluronidase (Hyaluronidase type IV-S) for 1 min. After removing the corona cells, only metaphase II oocytes were injected, and cultured in 30  $\mu\text{l}$  microdrops of ISM1™ (Medicult).

### Evaluation at day +1

At 16–18 h post-insemination, normal fertilization was confirmed by the presence of two pronuclei (PN) and two polar bodies (PB). All zygotes were then observed under the inverted microscope at  $\times 10$ , 20 or 40 magnification to determine the following morphological features:



**Figure 1** Pronucleus (PN) with nucleolar precursor body (NPB) aligned (A). Light halo.

- Comparative size of the male versus female PN categorized as equal or unequal.
- Degree of NPB organization categorized as aligned, semi-aligned or non-aligned with respect to the PN abuttal line. (Figs. 1–3).
- Presence of a light halo (Fig. 1), defined as visibly less dense cytoplasm in the outer cortical zone. This was best observed using a magnification of  $\times 10$ .
- Presence of a dark halo, defined as visibly denser and darker cytoplasm in the central zone, around the PNs. This too was best observed using a magnification of  $\times 10$ .
- Placement of PBs, categorised as adjacent or separate (Fig. 4).
- PB fragmentation categorized as fragmented if either of the PBs showed fragmentation or non-fragmented if neither PB showed fragmentation.

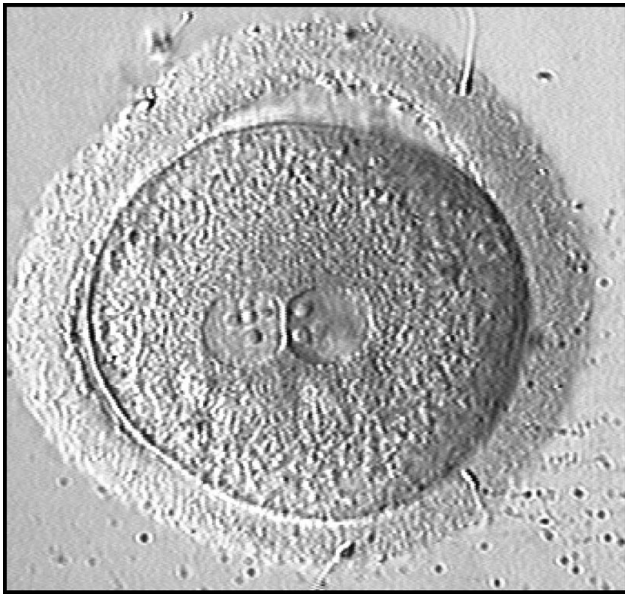
### Evaluation at day +2

At 48 h post-insemination, zygotes that had failed to cleave were identified and only these were included in this study.

We compared the blocked versus cleaved zygotes in terms of morphological features observed at 16–18 h post-insemination (Table 1).

### Statistical analysis

We used Windows SPSS (version 10.0), applying Pearson's chi-squared test, and Fisher's exact statistical test. A value of  $p < 0.05$  was considered as significant.



**Figure 2** Pronucleus (PN) with nucleolar precursor body (NPB), semi-aligned (S).

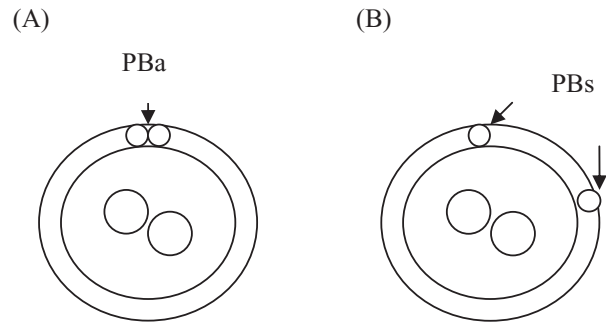


**Figure 3** Pronucleus (PN) with nucleolar precursor body (NPB), not aligned (NA).

## Results

Of 2105 zygotes, we found blockage in 43, which represents 2% of the total. This 2% was our reference point when evaluating the incidence of ZA for each morphological feature. Table 1 shows the comparison of blocked versus cleaved zygotes in terms of morphological features observed at 16–18 h post-insemination.

Three of these morphological parameters proved statistically significant with respect to the incidence of zygote arrest:



**Figure 4** Polar body (PB) placement. (A) PB adjacent (PBa); (B) PB separate (PBs).

**Table 1** Zygote morphological features and incidence of blockage/cleavage.

Morphological feature at 16–18 h	Blockage at 48 h, blocked vs. cleaved (%)	<i>p</i> -value
Equal sized PNs	28/1498(1.9)	0.505
Different sized PNs	13/556(2.3)	
1PN	2/51(3.9)	
Aligned NPB	7/690(1.0)	0.04 <sup>a</sup>
Semi-aligned NPB	12/575(2.1)	
Non-aligned NPB	24/840(2.9)	
Dark halo	24/1585(1.5)	0.004 <sup>a</sup>
No dark halo	19/520(3.7)	
Light halo	24/1640(1.5)	0.00 <sup>a</sup>
No light halo	19/465(4.1)	
PBs adjacent	33/1619(2.1)	0.55
PBs separate	10/486(2.1)	
No PB fragmentation	10/834(1.2)	0.084
PB fragmentation	33/1271(2.6)	

<sup>a</sup>Statistically significant ( $p < 0.05$ ).

NPB, nucleolar precursor bodies; PB, polar body; PN, pronucleus.

1. The degree of NPB organization. Thus, the lower the organizational degree, the higher the percentage of ZA: non-aligned (2.9%) > semi-aligned (2.1%) > aligned (1.0%),  $p < 0.05$ .
2. The presence of a light halo is a favorable morphological feature. Its absence significantly correlated with ZA: light halo (1.5%) < no light halo (4.1%),  $p = 0.001$ .
3. Similarly, the presence of a dark halo is favorable. This is usually observed together with the former feature. Its absence significantly correlated with ZA: dark halo (1.5%) < no dark halo (3.7%),  $p < 0.005$ .

With respect to the remaining features, in this series the comparative pronuclear size did not significantly correlate with ZA ( $p = 0.0505$ ) but it could be considered that unequal pronuclear sized zygotes had a tendency to have more ZA, and polar body placement or fragmentation showed no correlation with ZA (NS).



## Discussion

Fertilization reprogrammes the human egg and sperm from non-dividing, fully differentiated cells into totipotent cleaving blastomeres. Totipotency persists for three or four cell doublings, from the single cell zygote to the 8- to 16-cell morula. The stage of embryonic genome major activation is crucial for the success of preimplantation embryo development that means a correct maternal to embryo transition of gene expression (Kiessling, 2009).

Developmental arrest in embryos is a well known and much studied phenomenon (Chi, 2000; Hardy *et al.*, 2001; Racowsky, 2002; Favetta, 2007), however, failure of the zygote to cleave at all and thus become an embryo, is far less frequent and, consequently, has been little studied. What is known is that the *ZAR-1* gene may be considered as one of the transcriptional regulators acting during oocyte-to-embryo transition of gene expression (Uzbekova, 2006).

Also is a fact that the process that controls the centrosome and centriole duplication and the formation of a meiotic spindle during syngamy is distinct from that which takes place in somatic cells. If the centrosomes of both gametes become functional, mosaicism may be induced following the formation of abnormal meiotic spindles (Palermo *et al.*, 1994). In human reproduction, it is the sperm centrosome that controls the first mitotic division after fertilization. Functional failure of this centrosome may be responsible for failure of the first mitotic division.

From the molecular point of view, Rawe *et al.* (2003) have suggested that abnormal biogenesis or function of the zygotic centrosome could affect the recruitment of nuclear pore complexes and cytoplasm trafficking of annulate lamellae in some arrested zygotes, mainly in those showing pronuclear apposition.

Mammalian oocytes must undergo full cytoplasmic and nuclear maturation to at least metaphase II before they can develop viable embryos (Howlet & Bolton, 1985). Certain oocyte polypeptides are directly involved in embryogenesis by reorganizing the chromatin in preparation for DNA replication and activation of embryonic transcription. Premature insemination may interfere with these molecular processes, leading to ZA.

Chromosome disorder has been linked with early embryo development blockage and implantation failure and it is known that the vast majority of blocked zygotes and unfertilized oocytes show chromosome abnormalities (Benkhalifa *et al.*, 2003).

With respect to morphological features and ZA, zygotes whose pronuclei differed in size by more than 4  $\mu\text{m}$  have significantly increased frequency of DA (Sadowy *et al.*, 1998). One of the limitations of this study was that we did not measure differences

in diameter size: our data were qualitative, and all pronuclei pairs of different sizes were categorized as "different". Thus, we were unable to confirm this finding. However, we found a better developmental prognosis when pronuclei pairs were of equal size (NS). Of note, we not only included PNs of equal and different sizes but also those zygotes with a single PN, as more than 80% proceed from fertilized oocytes (Munne *et al.*, 1993), and in FISH studies concerning ploidy status of 1PN-derived embryos displayed a normal ploidy number of chromosomes in nearly half of them. Probably some monopronuclear stage can be mis-defined because of the clustering of some vacuolar or pronuclear membrane remnants and the cells have a 1PN appearance at the time of observation (Zhivkova, 2007). In circumstances of extremely limited availability of zygotes, we may be forced to use those with a single PN. Although not significant, we observed a marked tendency for 1PN zygotes to show higher rates of ZA (3.9 vs. 1.9 and 2.3% for equal and differently sized PNs respectively). Furthermore, Manor *et al.* (1999) found a higher incidence of chromosomal anomalies in embryos from zygotes with pronuclear dysmorphism, and even recommended applying preimplantation genetic diagnosis (PGD) in these cases prior to transfer.

Certain authors have related pronuclear morphology with a higher incidence of chromosomal anomalies (Gianaroli *et al.*, 2003). Others have developed a zygote morphological classification system showing correlation with improved implantation and pregnancy rates (Kattera & Chen, 2004). In addition to the above studies, Ebner *et al.* (2003) reported that the presence of a light halo in the zygote has a positive prognostic value on blastocyst quality. Also, Salumets *et al.* (2001) showed a positive correlation of good embryo morphology with light cytoplasmic halo and NPB polarization. These studies support our findings that the absence of these features is correlated with a higher incidence of ZA.

Concerning the organization of nucleolar precursor bodies, many studies confirm that when there exist polarization in both pronuclei in addition to be of similar size, there is improved embryo quality and development (Tesarik & Greco, 1999; Scott *et al.*, 2000; Gamiz *et al.*, 2003), and this conclusion is in accordance with our results that ZA is incremented when pronuclei are non-aligned, the non-polarized situation of nucleolar precursor bodies.

Also, the presence of a light halo in the zygote is associated with a higher rate of preimplantation development and implantation (Ebner *et al.*, 2003), and in accordance with this fact we observed that the absence of a light halo correlates better with ZA. In addition to the previous, we observed the presence or

absence of a dark halo as a separated morphological feature, considering that is localized in the central area of the zygote, surrounding the pronuclei as a dense cytoplasm area appearing as darker than the rest of the cytoplasm. The dark halo used to appear when light halo is present and is a consequence of cytoplasmic rearrangements (Payne, 1997). In our observations, the absence of a light halo and the absence of a dark halo correlate with a higher incidence of zygote arrest.

The factors underlying normal or abnormal embryo development are largely pre-determined at the 1-cell stage. Environmental factors influence these, but 'there will always be a group of embryos that will arrest and a group that will survive even under significantly substandard conditions' (Hardy *et al.*, 2001). Whatever the cause of ZA (inadequate oocyte maturation, environmental and molecular factors, or genetic-chromosomal abnormalities), a single simple static observation of pronuclear morphology allows identification of those zygotes with the highest probability for arresting at the zygote stage.

All these studies support the idea of not only basing embryo selection on embryo quality at day +2 or +3, but also considering morphological features at the zygote stage. At the same time, we may be able to obtain valuable information as to the developmental capacity of each zygote.

In certain circumstances, day-2 embryos are not feasible for religious/ethical considerations, and in such cases pronuclear morphology is the sole criterion for zygote transfer (Scott & Smith, 1998). Evaluation of morphology and knowledge of cleavage and implantation potential is of additional utility, especially for countries with legal restrictions regarding cryopreservation and embryo selection. In Germany for example, only three zygotes are allowed to be cultured for embryo transfer (Montag *et al.*, 2001).

To conclude, in this study developmental arrest observed at the first cleavage stage significantly correlated with three morphological features of the zygote: the absence of a light halo, the absence of a dark halo and non-aligned NPB. Given the need to improve our knowledge in zygote and embryo developmental potential to optimize the culture strategies and cryopreservation and to avoid multiple pregnancies, such information may contribute to suboptimal zygote detection.

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## References

- Benkhalifa, M., Kahraman, S., Caserta, D., Domez, E. & Qumsiyeh, M.B. (2003). Morphological and cytogenetic analysis of intact oocytes and blocked zygotes. *Prenat. Diagn.* **23**, 397–404.
- Chi, L., de Jesus, E., Adler, A., Grifo, J.A., Berkeley, A.S. & Krey, L.C. (2000). 2-cell stage arrest in human embryos: incidence and developmental potential in different culture media. *Fertil. Steril.* **74**(Suppl.1) S222.
- Ebner, T., Moser, M., Sommergruber, M., Gaiswinkler, U., Wiesinger, R., Puchner, M. & Tews, G. (2003). Presence, but not type or degree of extension, of a cytoplasmic halo has a significant influence on preimplantation development and implantation behaviour. *Hum. Reprod.* **18**, 2406–12.
- Favetta, L.A., St John, E.J., King, W.A. & Betts, D.H. (2007). High levels of p66<sup>shc</sup> and intracellular ROS in permanently arrested early embryos. *Free Radic Biol. Med.* **42**, 1201–10.
- Gámiz, P., Rubio, C., de los Santos, M.J., Mercader, A., Simón, C., Remohí, J. & Pellicer, A. (2003). The effect of pronuclear morphology on early development and chromosomal abnormalities in cleavage-stage embryos. *Hum. Reprod.* **18**, 2413–9.
- Gianaroli, L., Magli, M.C., Ferraretti, A.P., Fortini, D. & Griego, N. (2003). Pronuclear morphology and chromosomal abnormalities as scoring criteria for embryo selection. *Fertil. Steril.* **80**, 341–9.
- Hardy, K., Spanos, S., Becker, D., Iannelli, P., Winston, R.M.L. & Stark, J. (2001). From cell death to embryo arrest: mathematical models of human preimplantation embryo development. *Dev. Biol.* **98**, 1655–60.
- Howlet, S.K. & Bolton, V.N. (1985) Sequence and regulation of morphological and molecular events during the first cell cycle of mouse embryogenesis. *J. Embryol. Exp. Morph.* **87**, 175–206.
- Kattera, S. & Chen, C. (2004). Developmental potential of human pronuclear zygotes in relation to their pronuclear orientation. *Hum. Reprod.* **19**, 294–9.
- Kiessling, A., Bletsas, R., Desmarais, B., Mara, C., Kallianidis, K. & Loutradis, D. (2009). Evidence that human blastomere cleavage is under unique cell cycle control. *J. Assist. Reprod. Genet.* **26**, 187–95.
- Manor, D., Drugan, A., Stein, D., Pillar, M. & Itskovitz-Eldor, J. (1999). Unequal pronuclear size—a powerful predictor of embryonic chromosome anomalies. *J. Assist. Reprod. Genet.* **16**, 385–9.
- Montag, M., Van Der Ven, H. and [30 authors] on behalf of the German Pronuclear Morphology Study Group. (2001). Evaluation of pronuclear morphology as the only selection criterion for further embryo culture and transfer: results of a prospective multicentre study. *Hum. Reprod.* **16**, 2384–9.
- Munné, S., Tang, Y.X., Grifo, J. & Cohen, J. (1993). Origin of single pronucleated human zygotes. *J. Assist. Reprod. Genet.* **10**, 276–9.
- Palermo, G., Munné, S. & Cohen, J. (1994). The human zygote inherits its mitotic potential from the male gamete. *Hum. Reprod.* **9**, 1220–5.
- Payne, D., Flaherty, S.P., Barry, M.F. & Matthews, C.D. (1997). Preliminary observations on polar body extrusion and pronuclear formation in human oocytes using time-lapse video cinematography. *Hum. Reprod.* **12**, 532–41.

- Rawe, V.Y., Brugo Olmedo, S., Nodar, F.N., Poncio, R. & Sutovsky, P. (2003). Abnormal assembly of annulate lamellae and nuclear pore complexes coincides with fertilization arrest at the pronuclear stage of human zygotic development. *Hum. Reprod.* **18**, 576–82.
- Sadowy, S., Tomkin, G., Munné, S., Ferrara-Congedo, T. & Cohen, J. (1998). Impaired development of zygotes with uneven pronuclear size. *Zygote* **6**, 137–41.
- Sathananthan, A.H., Ratnam, S.S., Ng, S.C., Tarín, J.J., Gianaroli, L. & Trounson, A. (1996). The sperm centriole: its inheritance, replication and perpetuation in early human embryos. *Hum. Reprod.* **11**, 345–56.
- Salumets, A., Hydén-Granskog, C., Suikkari, A.M., Tiitinen, A. & Tuuri, T. (2001). The predictive value of pronuclear morphology of zygotes in the assessment of human embryo quality. *Hum. Reprod.* **16**, 2177–81.
- Scott, L. & Smith, S. (1998). The successful use of pronuclear embryo transfers the day following oocyte retrieval. *Hum. Reprod.* **13**, 1003–13.
- Scott, L., Alvero, R., Leondires, M. & Miller, B. (2000). The morphology of human pronuclear embryos is positively related to blastocyst development and implantation. *Hum. Reprod.* **15**, 2394–403.
- Tesarik, J. & Greco, E. (1999). The probability of abnormal preimplantation development can be predicted by a single static observation on pronuclear stage morphology. *Hum. Reprod.* **14**, 1318–23.
- Uzbekova, S., Roy-Sabau, M., Dalbiès-Tran, R., Perreau, C., Papillier, P., Mompert, F., Thelie, A., Pernetier, S., Cognie, J., Cadoret, V., Royere, D., Monget, P. & Mermillod, P. (2006). Zygote arrest I dene in pig, cattle and human: evidence of different transcript variants in male and female germ cells. *Reprod. Biol. Endocrinol.* **4**, 12.
- Wu, X., Viveiros, M.M., Eppig, J.J., Bai, Y., Fitzpatrick, S.L. & Matzuk, M.M. (2003). Zygote arrest 1 (*Zar 1*) is a novel maternal-effect gene critical for the oocyte-to-embryo transition. *Nat. Genet.* **33**, 187–91.
- Zhivkova, R.S., Delimitreva, S.H., Toncheva, D.I. & Vatev, I.T. (2007). Analysis of human unfertilized oocytes and pronuclear zygotes—correlation between chromosome/chromatin status and patient-related factors. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **130**, 73–83.