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# Bioinformatic approaches to draft the viral genome sequence of Canary Islands cases related to the multi-country Monkeypox virus 2022-outbreak

Adrián Muñoz-Barrera<sup>2#</sup>, Laura Ciuffreda<sup>3#</sup>, Julia Alcoba-Florez<sup>1#</sup>, Luis A. Rubio-Rodríguez<sup>2</sup>, Héctor Rodríguez-Pérez<sup>3</sup>, Helena Gil-Campesino<sup>1</sup>, Diego García-Martínez de Artola<sup>1</sup>, Josmar Salas-Hernández<sup>3</sup>, Julia Rodríguez-Núñez<sup>3</sup>, Antonio Íñigo-Campos<sup>2</sup>, Oscar Díez-Gil<sup>1</sup>, Rafaela González-Montelongo<sup>2</sup>, Agustín Valenzuela-Fernández<sup>4</sup>, José M. Lorenzo-Salazar<sup>2®</sup>, Carlos Flores<sup>2,3,5,6®\*</sup>

<sup>#</sup>Equal contribution as first authors

<sup>®</sup>Equal contribution

\*Corresponding author

<sup>1</sup>Servicio de Microbiología, Hospital Universitario Ntra. Sra. de Candelaria, 38010 Santa Cruz de Tenerife, Spain

<sup>2</sup>Genomics Division, Instituto Tecnológico y de Energías Renovables, 38600 Santa Cruz de Tenerife, Spain

 <sup>3</sup>Fundación Canaria Instituto de Investigación Sanitaria de Canarias at the Research Unit, Hospital Universitario Ntra. Sra. de Candelaria, 38010 Santa Cruz de Tenerife, Spain
<sup>4</sup>Laboratorio de Inmunología Celular y Viral, Unidad de Farmacología, Facultad de Medicina, Universidad de La Laguna, 38200 San Cristóbal de La Laguna, Spain

<sup>5</sup>CIBER de Enfermedades Respiratorias, Instituto de Salud Carlos III, 28029 Madrid, Spain <sup>6</sup>Facultad de Ciencias de la Salud, Universidad Fernando Pessoa Canarias, 35450 Las Palmas de Gran Canaria, Spain

# **Correspondence:**

Dr. Carlos Flores Unidad de Investigación Hospital Universitario Nuestra Señora de Candelaria Carretera del Rosario s/n 38010 Santa Cruz de Tenerife Spain Email: <u>cflores@ull.edu.es</u> Phone: (+34) 922 60 29 38

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### Abstract

During May 2022, the World Health Organization declared community transmission of monkeypox (MPXV) due to a multi-country outbreak. In Europe, several cases of this outbreak were detected in the Canary Islands (Spain). Here we describe the combination of DNA sequencing and bioinformatic approaches, including methods for *de novo* genome assembly and short- and long-read technologies, used to reconstruct the first MPXV genome isolated in the Canary Islands on the 31<sup>st</sup> of May 2022 from a male adult patient with mild symptoms. We obtained the best results using a reference-based approach with short reads, evidencing 46-67 nucleotide variants against sequences from the 2018-2019 outbreak, and placing the sequence in the B.1 clade. This study demonstrates the potential of metagenomics sequencing for rapid and precise pathogen identification.

## Keywords

Bioinformatics, de novo genome assembly, viral surveillance, monkeypox, MPXV

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### Introduction

Monkeypox virus (MPXV) is a zoonotic Orthopoxvirus (OPV) (family *Poxviridae*) [1,2], endemic to West and Central Africa [3,4]. MPXV has been described in humans in Central and Western Africa (occurring mainly in tropical forest areas of Central Africa) as well as in other parts of the world [5–10]. Around the 13<sup>th</sup> of May 2022, MPXV cases were reported in several countries and WHO declared community transmission of the virus [11]. Most reported cases so far have been presented through sexual health or other health services and have involved mainly men who have sex with men [12,13]. By early June 2022, 129 viral genomes had been deposited at GISAID [14] with 46 SNPs shared by all these sequences and differing from the viral genome sequences from the 2018-2019 MPXV outbreak [15]. Preliminary data from polymerase chain reaction (PCR) assays indicate that these MPXV strains detected in Europe and other non-endemic areas belong to the West African clade [16].

In Europe, several cases of MPXV infection have been associated with 2022 outbreaks in the Canary Islands and Spain [16]. A few viral sequences from samples collected in Spain have been reported [17]. As of June 2022, a total of 15 positive cases were confirmed in the Canary Islands [18], which have increased to a total of 176 by early November 2022. Here we describe the combination of methodological approaches to obtain the draft sequences of the first MPVX viral genome isolated in the Canary Islands on the 31<sup>st</sup> of May 2022 from a male adult patient with one week-onset mild symptoms (fever, odynophagia) and presenting at the Emergency Room but not necessitating hospital admission. An expedited description of the case and the resulting draft sequences was publicly posted in mid-June 2022 [19].

### **Materials and Methods**

### DNA extraction and PCR testing

Viral DNA was extracted at the Hospital Universitario Ntra. Sra. de Candelaria (Santa Cruz de Tenerife, Spain) from five samples (nasopharyngeal swab, lesion crust, and vesicles) from the same patient using the eMAG system (Biomerieux) following manufacturer's instructions. Virus inactivation was conducted under a biosafety class II cabinet (TELSTAR bio-II-A), following ECDC procedures [20]. Diagnosis of MPXV infection was confirmed using the LightMix Modular Orthopox (Roche) and a real-time PCR assay described elsewhere [21]. This assay yielded threshold cycle values in the range of 17 to 33 in these samples.

## Short- and long-read DNA sequencing

Five independent DNA dual index libraries (one for each sample) were processed at Instituto Tecnológico y de Energías Renovables (ITER) with Nextera XT DNA Library Preparation Kit (Illumina Inc.), following the manufacturer's recommendations with manual library normalization, and pooled prior to sequencing. The quality of the libraries was assessed with a D1000 ScreenTape kit on the 4200 TapeStation System (Agilent). Library concentrations ranged from 7.4 to 10.4 nM, and showed a fragmentation profile ranging from 721 to 808 bp. The mean fragment size for the sequencing pool was 677 bp as measured with a D1000 High Sensitivity ScreenTape kit (Agilent). Paired-end sequences were obtained on a MiSeq Sequencing System (Illumina Inc.), using the reagent kit v3 chemistry with 150 cycles and an expected throughput of 3.3-3.8 Gb. The pool concentration was 15 pM, and 5% of PhiX Control V3 was used as the internal control.

DNA libraries for nanopore sequencing were also prepared from the sample with the highest yield (taken from a skin lesion exudate) using the Rapid Barcoding kit (SQK-RBK004) from Oxford Nanopore Technologies (ONT). To increase the quantity of the starting material, the protocol used 30 to 45 ng of the DNA extract in 7.5  $\mu$ l for 12 independently barcoded libraries that were pooled in order to obtain the maximum yield from the run. The pooled libraries were loaded onto an R9.4.1 flow cell and were run in a MinION (ONT) for 42 hours. Basecalling of raw ONT signal data as well as demultiplexing and adapter trimming was carried out using Guppy v.6.0.7 with default parameters and the high-accuracy basecalling model.

## Bioinformatic analyses and assembly comparisons

As the first step, the individual demultiplexed FASTQ pair of Illumina files were interleaved with BBMap (Reformat tool) and then merged into a single interleaved FASTQ file. Then, two different bioinformatic tools were tested to identify and remove the human reads: Kraken 2 [22] and NCBI SRA Human Scrubber v.1.0.2021\_05\_05 (only used for Illumina sequencing data). The remaining Illumina and ONT reads were subjected to different bioinformatic procedures to obtain draft sequences (**Figure 1**).

On the one hand, a reference-based analysis was conducted with Illumina unclassified reads that were mapped to the MPVX viral genome MPXV-UK\_P2 (GenBank MT903344.1) by means of three alternative aligners: minimap2 v.2.24-r1122 [23], BWA-MEM v.0.7.17 [24], and Bowtie2 v.2.4.5 [25]. At this stage, duplicate metrics from PICARD v.2.18.7 [26], and coverage metrics from SAMtools v.1.6 [27] and mosdepth v.0.3.3 [28] were obtained from the remaining interleaved paired-end reads. Variant calling was carried out with two alternative algorithms: iVar v1.3.1 [29] and LoFreq v.2.1.5 [30] using default parameters against the MPXV MT903344.1 genome. For downstream analyses, a consensus sequence was obtained by piping a SAMtools v.1.6 pileup with iVar v.1.3.1 consensus as described elsewhere [31].

On the other hand, a hybrid *de novo* assembly was obtained by combining the filtered Illumina and ONT reads using custom script based on the Unicycler v.0.5.0 [32] assembler. Bandage v.0.9.0 [33] was used to visualize the resulting contigs in the assembly. A refined version of this hybrid *de novo* assembly was obtained after running Kraken 2 v.2.1.2 with PlusPF database to remove non-viral assembled contigs. The consensus sequence of this assembly was obtained by mapping resulting contigs to MPXV MT903344.1 genome and piping SAMtools v.1.6 pileup with BCFtools v.1.6 and seqtk v.1.3-r106.

Finally, the two selected consensus sequences (Illumina-only and hybrid *de novo* assembly) were compared against the MT903344.1 as the reference genome with QUAST v.5.0.2 [34].

## Phylogenetic analysis

The most complete consensus sequence resulting from the previous stage was aligned with 126 MPXV sequences downloaded from NCBI GenBank (**Table S1**) using MAFFT v.7.505 [35]. A phylogenetic analysis was performed using both IQ-TREE v.2.2.0.3 [36] with the K3Pu+F+I model as the best-predicted model and default parameters, and using a local instance of Nextstrain monkeypox [37].

## Results

The Illumina sequencing run produced 3.88 Gb and 25.5 M reads in total. A mapping of 101,814 and 100,897 Illumina reads was obtained using minimap2 on the NCBI SRA Human Scrubber (mean depth: 38.3x) and Kraken 2 (mean depth: 38.1x), respectively, thus providing equivalent results (Table 1). We estimated as few as 2.81% of duplicated reads and that 99% of the MPXV genome was covered  $\geq 1x$ , with a fraction of 85% of the viral genome covered at  $\geq 10x$ . The combination of mapper and variant caller that yielded the smallest and the largest number of nucleotide variants against the reference was BWA+LoFreq (46 nucleotide variants) and minimap2+iVar (67 nucleotide variants), respectively (Table 1). In order to maintain the maximum sequence variability for downstream analyses, the consensus sequence for Illumina-only reads was obtained for the minimap2+iVar combination (total size of 197,221 bp), providing a near-fully complete viral genome (99.91%) against the reference.

The ONT run provided 1.98 Gb and a total of 1.38 M reads, ranging from 499 to 101,895 bp in length, with a mean length of 1,432 bases. ONT sequencing provided 2,246 non-human mapping reads after filtering with Kraken 2, thus, a theoretical viral genome depth of 14.9x. A hybrid *de novo* assembly based on Illumina and ONT Kraken 2-filtered reads was performed and resulted in four contigs (**Figure 2**). Contigs 1 and 2 accounted for 186,315 bp and 4,703 bp (191,018 bp total), respectively, and mapped to Monkeypox virus Zaire-96-I-16. Contigs 3 and 4 spanned 10,530 bp in total but did not map to the MPXV reference. Thus, a consensus sequence was built from this hybrid *de novo* assembly (including only contigs 1 and 2) and the MT903344.1 MPXV reference genome, spanning a total size of 197,222 bp. Note, however, that given that this sequence includes 6,471 undetermined bases, the consensus sequence only covered 96.75% of the reference genome (**Table 2**).

Besides the number of undetermined bases, which were much more in the hybrid *de novo* assembly sequence than in the Illumina-only consensus sequence, we observed small differences overall between the two MPXV genome assemblies obtained (**Table 2**). In brief, the hybrid *de novo* assembly was able to retrieve a slightly lower proportion of the reference genome, although showing a similar GC content and having fewer mismatches than the Illumina-only assembly.

Based on the above findings supporting more completeness for the Illumina-only assembly, we opted for placing this consensus sequence in phylogenetic context (**Figure 3**). This analysis indicated that the draft MPXV genome sequence belongs to the so-called West African clade or B.1 [38,39]. In addition, the closest sequences were related to the Slovenian-MPXV GenBank-released genomes, contributing further evidence of community spread in the present worldwide MPXV outbreak.

#### Discussion

Here we provide the draft sequences of the first MPVX viral genome isolated in the Canary Islands on 31 May 2022, corresponding to the B.1 clade that was observed across Europe and other non-endemic areas during the outbreak. For that, we have used diverse sequencing and bioinformatic approaches, including methods for *de novo* genome assembly and combining short and long sequencing reads. The best results (higher sequence similarity and higher genome coverage compared to the reference) were obtained using a reference-based approach with Illumina-only reads. With this approximation, and using combinations of mappers and variant callers, between 46 and 67 nucleotide variants were

observed when compared to the reference viral genome, which is compatible with the divergence of this new MPXV outbreak from that of 2018-2019 [15].

For this first characterization, we have relied on a metagenomic sequencing approach, providing a low proportion of sequence reads corresponding to the viral genome. In the study, only 0.39% and 0.16% of the reads obtained from the Illumina and ONT runs, respectively, mapped to the MPXV reference. This approach is straightforward and is not as dependent on the viral sequence rearrangements in the outbreaks, as is common for OPVs, demonstrating its value for detecting RNA and DNA viral pathogens in a few hours [40]. However, the costs per sample as well as the viral DNA concentration requirements in the patient samples hinders an extended use. After this work was completed [19], a more sensitive approach based on tiling amplicon sequencing both for Illumina and ONT workflows was developed and validated across a number of laboratories for MPXV [41]. Despite this method being more cost-efficient, a periodic long-read metagenomics sequencing was recommended by the authors to monitor the emergence of viral variants with genomic rearrangements.

Overall, our results provide a proof of concept of the potential of introducing sequencing technologies, and metagenomics in particular, for rapid and precise pathogen diagnosis and surveillance.

#### Ethics statement

The study was conducted at the University Hospital Nuestra Señora de Candelaria (Santa Cruz de Tenerife, Spain) during May 2022. The institutional review board approved the study (ethics approval number: CHUNSC\_2022\_83).

#### Data availability statement

The code used for this study is available at <u>https://github.com/genomicsITER/monkeypox</u>. The Illumina-only and the hybrid *de novo* assembly consensus MPXV sequences have been released in the NCBI GenBank with accessions ON782054 and ON782055, respectively.

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### **Conflicts of Interest**

The authors declare no conflict of interest.

### **Author contributions**

JAF, JMLS and CF designed the study. AMB, LC, JAF, LRR, HRP, RGM, JSH, JRN, AIC, DGM, HGC, ODG, and CF participated in data acquisition. AMB, LRR, HRP, LC, JAF, JMLS

and CF performed the analyses and data interpretation. LC, AVF, JMLS and CF wrote the draft of the manuscript. CF obtained funding. All authors contributed in the critical revision and final approval of the manuscript.

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## References

- Esposito J, Fenner F, Others. Poxviruses. Fields Virology (4th ed). Lippincott Williams & Wilkins; 2001.
- Ryan KJ, George Ray C, Sherris JC. Sherris Medical Microbiology: An Introduction to Infectious Diseases. McGraw-Hill; 2004.
- 3. Ligon BL. Monkeypox: a review of the history and emergence in the Western hemisphere. Semin Pediatr Infect Dis. 2004;15: 280–287.
- 4. Jezek Z, Fenner F. Human Monkeypox. Melnick JL, editor. Basel, Karger; 1988. pp. 1– 140.
- 5. Parker S, Nuara A, Buller RML, Schultz DA. Human monkeypox: an emerging zoonotic disease. Future Microbiol. 2007;2: 17–34.
- Gispen R, Brand-Saathof BB, Hekker AC. Monkeypox-specific antibodies in human and simian sera from the Ivory Coast and Nigeria. Bull World Health Organ. 1976;53: 355– 360.
- Hutin YJF, Williams RJ, Malfait P, Pebody R, Loparev V, Ropp S, et al. Outbreak of Human Monkeypox, Democratic Republic of Congo, 1996 to 1997. Emerging Infectious Disease journal. 2001;7: 434.
- 8. Di Giulio DB, Eckburg PB. Human monkeypox: an emerging zoonosis. Lancet Infect Dis. 2004;4: 15–25.
- 9. Parker S, Buller RM. A review of experimental and natural infections of animals with monkeypox virus between 1958 and 2012. Future Virol. 2013;8: 129–157.
- Reed KD, Melski JW, Graham MB, Regnery RL, Sotir MJ, Wegner MV, et al. The detection of monkeypox in humans in the Western Hemisphere. N Engl J Med. 2004;350: 342–350.
- 11. Multi-country monkeypox outbreak in non-endemic countries. [cited 7 Nov 2022]. Available: https://www.who.int/emergencies/disease-outbreak-news/item/2022-DON385
- Multi-country monkeypox outbreak in non-endemic countries: Update. [cited 7 Nov 2022]. Available: https://www.who.int/emergencies/disease-outbreak-news/item/2022-DON388
- 13. Multi-country monkeypox outbreak: situation update. [cited 7 Nov 2022]. Available: https://www.who.int/emergencies/disease-outbreak-news/item/2022-DON390
- 14. Shu Y, McCauley J. GISAID: Global initiative on sharing all influenza data from vision to reality. Euro Surveill. 2017;22. doi:10.2807/1560-7917.ES.2017.22.13.30494
- Multi-country outbreak of Monkeypox virus: genetic divergence and first signs of microevolution. In: Virological [Internet]. 23 May 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/multi-country-outbreak-of-monkeypox-virus-genetic-divergenceand-first-signs-of-microevolution/806
- 16. Illumina whole-genome sequence of Monkeypox virus in a patient travelling from the Canary Islands to France. In: Virological [Internet]. 28 May 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/illumina-whole-genome-sequence-of-monkeypox-virus-in-a-patient-travelling-from-the-canary-islands-to-france/829
- 17. UPDATE: Two draft genomes from Madrid, Spain, of the Monkeypox virus 2022 outbreak. In: Virological [Internet]. 8 Jun 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/update-two-draft-genomes-from-madrid-spain-of-the-monkeypox-virus-2022-outbreak/848
- 18. Health related news from the Government of the Canary Islands. [cited 7 Nov 2022]. Available: https://www3.gobiernodecanarias.org/noticias/sanidad-contabiliza-tres-casos-

confirmados-y-tres-en-estudio-de-viruela-del-mono-desde-el-viernes/

- A draft of the first genome sequence of Monkeypox virus associated with the multicountry outbreak in May 2022 from the Canary Islands, Spain. In: Virological [Internet].
  Jun 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/a-draft-of-the-firstgenome-sequence-of-monkeypox-virus-associated-with-the-multi-country-outbreak-inmay-2022-from-the-canary-islands-spain/864
- 20. ECDC, Factsheet for health professionals on monkeypox. In: European Centre for Disease Prevention and Control [Internet]. [cited 7 Nov 2022]. Available: https://www.ecdc.europa.eu/en/all-topics-z/monkeypox/factsheet-health-professionals
- Li Y, Zhao H, Wilkins K, Hughes C, Damon IK. Real-time PCR assays for the specific detection of monkeypox virus West African and Congo Basin strain DNA. J Virol Methods. 2010;169: 223–227.
- 22. Wood DE, Lu J, Langmead B. Improved metagenomic analysis with Kraken 2. Genome Biol. 2019;20: 257.
- 23. Li H. Minimap2: pairwise alignment for nucleotide sequences. Bioinformatics. 2018;34: 3094–3100.
- 24. Li H. Aligning sequence reads, clone sequences and assembly con\*gs with BWA-MEM. figshare; 2014. doi:10.6084/M9.FIGSHARE.963153.V1
- 25. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. Nat Methods. 2012;9: 357–359.
- 26. Broad Institute. Picard Toolkit. 2018 [cited 7 Nov 2022]. Available: http://broadinstitute.github.io/picard/
- 27. Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, et al. Twelve years of SAMtools and BCFtools. Gigascience. 2021;10. doi:10.1093/gigascience/giab008
- 28. Pedersen BS, Quinlan AR. Mosdepth: quick coverage calculation for genomes and exomes. Bioinformatics. 2018;34: 867–868.
- Grubaugh ND, Gangavarapu K, Quick J, Matteson NL, De Jesus JG, Main BJ, et al. An amplicon-based sequencing framework for accurately measuring intrahost virus diversity using PrimalSeq and iVar. Genome Biol. 2019;20: 8.
- 30. Wilm A, Aw PPK, Bertrand D, Yeo GHT, Ong SH, Wong CH, et al. LoFreq: a sequencequality aware, ultra-sensitive variant caller for uncovering cell-population heterogeneity from high-throughput sequencing datasets. Nucleic Acids Res. 2012;40: 11189–11201.
- First French draft genome sequence of Monkeypox virus, may 2022. In: Virological [Internet]. 26 May 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/first-frenchdraft-genome-sequence-of-monkeypox-virus-may-2022/819
- 32. Wick RR, Judd LM, Gorrie CL, Holt KE. Unicycler: Resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol. 2017;13: e1005595.
- 33. Wick RR, Schultz MB, Zobel J, Holt KE. Bandage: interactive visualization of de novo genome assemblies. Bioinformatics. 2015;31: 3350–3352.
- 34. Gurevich A, Saveliev V, Vyahhi N, Tesler G. QUAST: quality assessment tool for genome assemblies. Bioinformatics. 2013;29: 1072–1075.
- 35. Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 2013;30: 772–780.
- 36. Nguyen L-T, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. Mol Biol Evol. 2015;32: 268–274.
- 37. Hadfield J, Megill C, Bell SM, Huddleston J, Potter B, Callender C, et al. Nextstrain: realtime tracking of pathogen evolution. Bioinformatics. 2018;34: 4121–4123.
- Urgent need for a non-discriminatory and non-stigmatizing nomenclature for monkeypox virus. In: Virological [Internet]. 16 Aug 2022 [cited 7 Nov 2022]. Available: https://virological.org/t/urgent-need-for-a-non-discriminatory-and-non-stigmatizingnomenclature-for-monkeypox-virus/853
- 39. Cohen J. Rename monkeypox strains to remove geographic stigma, researchers say. 2022. doi:10.1126/science.add4325

- 40. Alcolea-Medina A, Charalampous T, Snell LB, Aydin A, Alder C, Maloney G, et al. Novel, Rapid Metagenomic Method to Detect Emerging Viral Pathogens Applied to Human Monkeypox Infections. 2022. doi:10.2139/ssrn.4132526
- 41. Chen NFG, Chaguza C, Gagne L, Doucette M, Smole S, Buzby E, et al. Multi-site validation of an amplicon-based sequencing approach for human monkeypox virus. medRxiv. 2022. doi:10.1101/2022.10.14.22280783

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**Table 1.** Comparative of mapped Illumina reads and coverage using different aligners (Bowtie2, BWA, Minimap2) and called variants using iVar and LoFreq callers.

MPXV01	Kraken2							
Total reads	Non-human Reads	Aligners	Mapped reads	Coverage	iVar	LoFreq		
		Bowtie2	101,092 (2.52%)	38.15	65	47		
	4,009,480	BWA	101,456 (2.53%)	38.25	56	46		
		Minimap2	100,907 (2.52%)	38.08	67	48		
	NCBI SRA Human Scrubber							
51,042,414								
	Non-human Reads	Aligners	Mapped reads	Coverage	iVar	LoFreq		
	Non-human Reads	Aligners Bowtie2	Mapped reads 100,851 (0.96%)	Coverage 38.34	iVar 54	LoFreq 30		
	Non-human Reads	Aligners Bowtie2 BWA	Mapped reads       100,851 (0.96%)       105,484 (1.00%)	Coverage       38.34       38.84	iVar 54 58	LoFreq 30 46		

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Metrics	Hybrid <i>de nov</i> o assembly (ON782054)	Illumina-only (ON782055)
# contigs	1	1
Largest contig (bp)	197,222	197,221
Total length (bp)	197,222	197,221
Reference length (bp)	197,233	197,233
GC (%)	32.93	33.02
Reference GC (%)	33.02	33.02
N50	197,222	197,221
L50	1	1
# misassemblies	0	0
Genome fraction (%)	96.75	100.00
Duplication ratio	1.034	1.000
# N's per 100 kbp	3,281.07	92.79
# mismatches per 100 kbp	22.01	25.86
# indels per 100 kbp	0.00	1.52
Largest alignment (bp)	190,825	197,221
Total aligned length (bp)	190,825	197,221
NA50	190,825	197,221
LA50	1	1

## Table 2. Assessment of the two MPXV genome assemblies against the MPXV reference.



**Figure 1.** Full bioinformatic pipeline to obtain the MPXV sequences from Illumina-read only and the hybrid *de novo* assembly to infer phylogenetic relationships with other MPXV viral genomes available from public repositories.



Figure 2. Bandage representation of the hybrid *de novo* assembly based on long-read sequencing technology.





**Figure 3.** A phylogenetic tree depicting the draft MPXV sequence isolated on May 31, 2022 from a patient from the Canary Islands along with publicly available sequences at the NCBI GenBank.

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## **Supplementary Material**

Accession	Country	Collection date	Reference	Length (bp)	NCBI GenBank
MG693724.1	Nigeria	2017-12-31	Faye et al. 2017	183,055	https://www.ncbi.nlm.nih.g ov/nuccore/MG693724.1
MT903338.1	Nigeria	2018-01-01	Mauldin et al. 2020	197,559	https://www.ncbi.nlm.nih.g ov/nuccore/MT903338.1
NC_063383.1	Nigeria	2018-08-01	Mauldin et al. 2022	197,209	https://www.ncbi.nlm.nih.g ov/nuccore/NC 063383.1
MT903340.1	Nigeria	2018-07-31	Mauldin et al. 2020	197,209	https://www.ncbi.nlm.nih.g ov/nuccore/MT903340.1
MK783030.1	Nigeria	2017-11-30	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783030.1
MT903337.1	Nigeria	2018-01-22	Mauldin et al. 2020	197,547	https://www.ncbi.nlm.nih.g ov/nuccore/MT903337.1
MT903339.1	Nigeria	2018-01-01	Mauldin et al. 2020	197,556	https://www.ncbi.nlm.nih.g ov/nuccore/MT903339.1
MK783033.1	Nigeria	2017-10-09	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783033.1
MG693723.1	Nigeria	2017-12-31	Faye et al. 2017	182,482	https://www.ncbi.nlm.nih.g ov/nuccore/MG693723.1
MK783032.1	Nigeria	2017-11-30	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783032.1
ON675438.1	USA	2022-05-01	Gigante et al. 2022	196,919	https://www.ncbi.nlm.nih.g ov/nuccore/ON675438.1
ON676707.1	USA	2021-07-24	Gigante et al. 2022	197,265	https://www.ncbi.nlm.nih.g ov/nuccore/ON676707.1
ON674051.1	USA	2022-05-01	Gigante et al. 2022	197,166	https://www.ncbi.nlm.nih.g ov/nuccore/ON674051.1
MK783029.1	Nigeria	2017-12-06	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783029.1
MK783027.1	Nigeria	2017-11-09	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783027.1
MK783028.1	Nigeria	2017-11-09	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783028.1

Table S1. Acknowledgement and accession numbers used for phylogenetic analysis.

MK783031.1	Nigeria	2017-11-09	Yinka- Ogunleye et al. 2019	197,566	https://www.ncbi.nlm.nih.g ov/nuccore/MK783031.1
MN648051.1	Israel	2018-10-04	Cohen Gihon et al. 2019	197,417	https://www.ncbi.nlm.nih.g ov/nuccore/MN648051.1
MT250197.1	Singapor e	2019-01-01	Yong et al. 2020	197,551	https://www.ncbi.nlm.nih.g ov/nuccore/MT250197.1
MT903341.1	Nigeria	2018-08-14	Mauldin et al. 2020	185,309	https://www.ncbi.nlm.nih.g ov/nuccore/MT903341.1
ON676708.1	USA	2021-11-01	Gigante et al. 2022	197,173	https://www.ncbi.nlm.nih.g ov/nuccore/ON676708.1
ON585036.1	Portugal	2022-05-15	Isidro et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON585036.1
ON751962.1	Brazil	2022-06-07	Claro et al. 2022	197,420	https://www.ncbi.nlm.nih.g ov/nuccore/ON751962.1
ON645312.1	United Kingdom	2022-05-11	Alcolea-Medina et al. 2022	184,777	https://www.ncbi.nlm.nih.g ov/nuccore/ON645312.1
ON694335.1	German y	2022-05-23	Brinkmann et al. 2022	196,400	https://www.ncbi.nlm.nih.g ov/nuccore/ON694335.1
ON682263.3	German y	2022-05-04	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682263.3
ON619838.2	United Kingdom	2022-05-23	Groves et al. 2022	197,205	https://www.ncbi.nlm.nih.g ov/nuccore/ON619838.2
ON619837.2	United Kingdom	2022-05-23	Groves et al. 2022	197,181	https://www.ncbi.nlm.nih.g ov/nuccore/ON619837.2
ON585030.1	Portugal	2022-05-15	Isidro et al. 2022	197,208	https://www.ncbi.nlm.nih.g ov/nuccore/ON585030.1
ON676704.1	USA	2022-05-23	Gigante et al. 2022	197,176	https://www.ncbi.nlm.nih.g ov/nuccore/ON676704.1
ON585029.1	Portugal	2022-05-04	Isidro et al. 2022	197,068	https://www.ncbi.nlm.nih.g ov/nuccore/ON585029.1
ON676706.1	USA	2022-05-01	Gigante et al. 2022	196,996	https://www.ncbi.nlm.nih.g ov/nuccore/ON676706.1
ON631963.1	Australia	2022-05-04	Hammerschlag et al. 2022	197,443	https://www.ncbi.nlm.nih.g ov/nuccore/ON631963.1
ON649717.1	Portugal	2022-05-21	Isidro et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON649717.1
ON720848.1	Spain	2022-05-27	Buenestado- Serrano et al. 2022	197,044	https://www.ncbi.nlm.nih.g ov/nuccore/ON720848.1
ON755235.1	German	2022-06-01	Brinkmann et	197,140	https://www.ncbi.nlm.nih.g

	у		al. 2022		ov/nuccore/ON755235.1
ON754989.1	Canada	2022-06-06	Croxen et al. 2022	197,264	https://www.ncbi.nlm.nih.g ov/nuccore/ON754989.1
ON622720.1	Switzerla nd	2022-05-24	Laubscher et al. 2022	197,128	https://www.ncbi.nlm.nih.g ov/nuccore/ON622720.1
ON745225.1	Spain	2022-05-27	Buenestado- Serrano et al. 2022	197,205	https://www.ncbi.nlm.nih.g ov/nuccore/ON745225.1
ON649724.1	Portugal	2022-05-23	lsidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649724.1
ON649719.1	Portugal	2022-05-23	lsidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649719.1
ON631241.1	Slovenia	2022-05-24	Zakotnik et al. 2022	197,520	https://www.ncbi.nlm.nih.g ov/nuccore/ON631241.1
ON649721.1	Portugal	2022-05-19	Isidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649721.1
ON649718.1	Portugal	2022-05-20	Isidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649718.1
ON649723.1	Portugal	2022-05-20	lsidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649723.1
ON649722.1	Portugal	2022-05-19	Isidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649722.1
ON649725.1	Portugal	2022-05-23	Isidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649725.1
ON563414.3	USA	2022-05-01	Gigante et al. 2022	197,205	https://www.ncbi.nlm.nih.g ov/nuccore/ON563414.3
ON755040.1	France	2022-05-20	Jarjava et al. 2022	196,947	https://www.ncbi.nlm.nih.g ov/nuccore/ON755040.1
ON649710.1	Portugal	2022-05-18	Isidro et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON649710.1
ON649715.1	Portugal	2022-05-20	Isidro et al. 2022	197,191	https://www.ncbi.nlm.nih.g ov/nuccore/ON649715.1
ON614676.1	Italy	2022-05-18	Gruber et al. 2022	190,289	https://www.ncbi.nlm.nih.g ov/nuccore/ON614676.1
ON595760.2	Switzerla nd	2022-05-19	Laubscher et al. 2022	197,056	https://www.ncbi.nlm.nih.g ov/nuccore/ON595760.2
ON755253.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755253.1
ON585035.1	Portugal	2022-05-15	lsidro et al. 2022	197,220	https://www.ncbi.nlm.nih.g ov/nuccore/ON585035.1

ON736420.1	Canada	2022-05-31	Croxen et al. 2022	197,201	https://www.ncbi.nlm.nih.g ov/nuccore/ON736420.1
ON755243.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755243.1
ON585034.1	Portugal	2022-05-15	lsidro et al. 2022	197,208	https://www.ncbi.nlm.nih.g ov/nuccore/ON585034.1
ON585031.1	Portugal	2022-05-15	Isidro et al. 2022	197,208	https://www.ncbi.nlm.nih.g ov/nuccore/ON585031.1
ON755234.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755234.1
ON585032.1	Portugal	2022-05-17	Isidro et al. 2022	197,211	https://www.ncbi.nlm.nih.g ov/nuccore/ON585032.1
ON755231.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755231.1
ON755237.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755237.1
ON755241.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755241.1
ON676703.1	USA	2022-05-01	Gigante et al. 2022	197,096	https://www.ncbi.nlm.nih.g ov/nuccore/ON676703.1
ON649879.1	Israel	2022-05-20	Israeli et al. 2022	196,753	https://www.ncbi.nlm.nih.g ov/nuccore/ON649879.1
ON649716.1	Portugal	2022-05-23	Isidro et al. 2022	197,193	https://www.ncbi.nlm.nih.g ov/nuccore/ON649716.1
ON755245.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755245.1
ON755246.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755246.1
ON682267.2	German y	2022-05-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682267.2
ON694341.1	German y	2022-05-04	Brinkmann et al. 2022	197,180	https://www.ncbi.nlm.nih.g ov/nuccore/ON694341.1
ON755247.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755247.1
ON644344.1	Italy	2022-05-25	Licastro et al. 2022	197,417	https://www.ncbi.nlm.nih.g ov/nuccore/ON644344.1
ON694336.1	German y	2022-05-01	Brinkmann et al. 2022	197,303	https://www.ncbi.nlm.nih.g ov/nuccore/ON694336.1
ON622713.1	Belgium	2022-05-22	Wawina et al. 2022	198,016	https://www.ncbi.nlm.nih.g ov/nuccore/ON622713.1

ON627808.1	USA	2022-05-20	Young et al. 2022	197,114	https://www.ncbi.nlm.nih.g ov/nuccore/ON627808.1
ON568298.1	German y	2022-05-19	Antwerpen et al. 2022	197,378	https://www.ncbi.nlm.nih.g ov/nuccore/ON568298.1
ON745215.1	Italy	2022-05-19	Giombini et al. 2022	197,213	https://www.ncbi.nlm.nih.g ov/nuccore/ON745215.1
ON682270.2	German y	2022-05-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682270.2
ON755039.1	France	2022-05-19	Jarjaval et al. 2022	196,172	https://www.ncbi.nlm.nih.g ov/nuccore/ON755039.1
ON649720.1	Portugal	2022-05-19	Isidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649720.1
ON649708.1	Portugal	2022-05-19	lsidro et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON649708.1
ON649712.1	Portugal	2022-05-19	Isidro et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON649712.1
ON682269.3	German y	2022-05-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682269.3
ON622712.1	Belgium	2022-05-19	Vanmechelen et al. 2022	198,010	https://www.ncbi.nlm.nih.g ov/nuccore/ON622712.1
ON676705.1	USA	2022-05-04	Gigante et al. 2022	197,154	https://www.ncbi.nlm.nih.g ov/nuccore/ON676705.1
ON585033.1	Portugal	2022-05-15	Isidro et al. 2022	197,220	https://www.ncbi.nlm.nih.g ov/nuccore/ON585033.1
ON649709.1	Portugal	2022-05-18	lsidro et al. 2022	197,204	https://www.ncbi.nlm.nih.g ov/nuccore/ON649709.1
ON649711.1	Portugal	2022-05-18	Isidro et al. 2022	197,185	https://www.ncbi.nlm.nih.g ov/nuccore/ON649711.1
ON615424.1	Netherla nds	2022-05-23	Oude Munnink et al. 2022	196,526	https://www.ncbi.nlm.nih.g ov/nuccore/ON615424.1
ON619835.2	United Kingdom	2022-05-01	Groves et al. 2022	197,205	https://www.ncbi.nlm.nih.g ov/nuccore/ON619835.2
ON619836.2	United Kingdom	2022-05-04	Groves et al. 2022	197,205	https://www.ncbi.nlm.nih.g ov/nuccore/ON619836.2
ON755239.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755239.1
ON755244.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755244.1
ON755249.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755249.1

ON755255.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755255.1
ON694330.1	German y	2022-05-31	Brinkmann et al. 2022	197,198	https://www.ncbi.nlm.nih.g ov/nuccore/ON694330.1
ON694342.1	German y	2022-05-17	Brinkmann et al. 2022	197,180	https://www.ncbi.nlm.nih.g ov/nuccore/ON694342.1
ON682266.2	German y	2022-05-04	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682266.2
ON682268.2	German y	2022-05-04	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682268.2
ON682264.3	German y	2022-05-31	Brinkmann et al. 2022	197,132	https://www.ncbi.nlm.nih.g ov/nuccore/ON682264.3
ON622718.1	Spain	2022-05-20	Martinez- Puchol et al. 2022	196,436	https://www.ncbi.nlm.nih.g ov/nuccore/ON622718.1
ON720849.1	Spain	2022-05-27	Buenestado- Serrano et al. 2022	197,096	https://www.ncbi.nlm.nih.g ov/nuccore/ON720849.1
ON755233.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755233.1
ON637939.1	German y	2022-05-31	Brinkmann et al. 2022	197,131	https://www.ncbi.nlm.nih.g ov/nuccore/ON637939.1
ON755254.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755254.1
ON585037.1	Portugal	2022-05-15	lsidro et al. 2022	196,295	https://www.ncbi.nlm.nih.g ov/nuccore/ON585037.1
ON585038.1	Portugal	2022-05-15	lsidro et al. 2022	196,305	https://www.ncbi.nlm.nih.g ov/nuccore/ON585038.1
ON649713.1	Portugal	2022-05-19	Isidro et al. 2022	196,291	https://www.ncbi.nlm.nih.g ov/nuccore/ON649713.1
ON649714.1	Portugal	2022-05-20	Isidro et al. 2022	197,193	https://www.ncbi.nlm.nih.g ov/nuccore/ON649714.1
ON755248.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755248.1
ON622722.2	France	2022-05-22		197,120	https://www.ncbi.nlm.nih.g ov/nuccore/ON622722.2
ON755251.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755251.1
ON755256.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755256.1
ON754984.1	Slovenia	2022-06-01	Zakotnik et al.	197,614	https://www.ncbi.nlm.nih.g

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ON754986.1	Slovenia	2022-06-01	Zakotnik et al. 2022	197,611	https://www.ncbi.nlm.nih.g ov/nuccore/ON754986.1
ON754985.1	Slovenia	2022-06-01	Zakotnik et al. 2022	197,650	https://www.ncbi.nlm.nih.g ov/nuccore/ON754985.1
ON609725.2	Slovenia	2022-05-23	Zakotnik et al. 2022	197,520	https://www.ncbi.nlm.nih.g ov/nuccore/ON609725.2
ON754987.1	Slovenia	2022-05-23	Zakotnik et al. 2022	197,653	https://www.ncbi.nlm.nih.g ov/nuccore/ON754987.1
ON682265.3	German y	2022-05-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON682265.3
ON755238.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755238.1
ON694332.1	German y	2022-05-31	Brinkmann et al. 2022	197,090	https://www.ncbi.nlm.nih.g ov/nuccore/ON694332.1
ON755236.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755236.1
ON694333.1	German y	2022-05-15	Brinkmann et al. 2022	197,210	https://www.ncbi.nlm.nih.g ov/nuccore/ON694333.1
ON755250.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755250.1
ON755242.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755242.1
ON694334.1	German y	2022-05-31	Brinkmann et al. 2022	197,236	https://www.ncbi.nlm.nih.g ov/nuccore/ON694334.1
ON694329.1	German y	2022-05-15	Brinkmann et al. 2022	197,271	https://www.ncbi.nlm.nih.g ov/nuccore/ON694329.1
ON694338.1	German y	2022-05-15	Brinkmann et al. 2022	197,434	https://www.ncbi.nlm.nih.g ov/nuccore/ON694338.1
ON755252.1	German y	2022-06-15	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755252.1
ON694337.1	German y	2022-05-31	Brinkmann et al. 2022	197,340	https://www.ncbi.nlm.nih.g ov/nuccore/ON694337.1
ON694340.1	German y	2022-05-31	Brinkmann et al. 2022	197,347	https://www.ncbi.nlm.nih.g ov/nuccore/ON694340.1
ON694331.1	German y	2022-05-18	Brinkmann et al. 2022	197,436	https://www.ncbi.nlm.nih.g ov/nuccore/ON694331.1
ON755232.1	German y	2022-06-01	Brinkmann et al. 2022	197,140	https://www.ncbi.nlm.nih.g ov/nuccore/ON755232.1
ON755240.1	German	2022-06-15	Brinkmann et	197,140	https://www.ncbi.nlm.nih.g

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ON637938.1	German y	2022-05-18	Brinkmann et al. 2022	197,131	https://www.ncbi.nlm.nih.g ov/nuccore/ON637938.1
ON694339.1	German y	2022-05-18	Brinkmann et al. 2022	197,415	https://www.ncbi.nlm.nih.g ov/nuccore/ON694339.1